

Species Composition of Ground Dwelling Staphylinid (*Coleoptera: Staphylinidae*) Communities in Apple and Pear Orchards in Hungary

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Species richness and composition of *Staphylinidae* communities were investigated at ground level when differently treated with pesticides and in abandoned apple and pear orchards in Hungary. Altogether 6099 individuals were collected belonging to 241 staphylinid species. 233 have been identified to species level and 8 staphylinid taxa were determined up to generic level.

More than 20% of the Hungarian staphylinid fauna was represented in the orchards. The similarity (Jaccard index) between apple and pear orchards at ground level were 54%. The species richness in each orchard varied between 23 and 100 species.

The most widely occurring species in orchard ground level were: *Dinaraea angustula*, *Palporus nitidulus*, *Tachyporus hypnorum*, *Sphenoma abdominale*, *Omalium caesum*, *Philonthus carbonarius*, *Drusilla canaliculata*, *Sepedophilus marshami*, *Mocyta orbata*, *Coprochara bipustulata*, *Mocyta fungi*, *Hyponygrus angustatus*, *Purrolinus laeticeps*, *Paraphallus linearis*, *Omalium cursor*, *Heterothops dissimilis* and *Atheta crassicornis*.

Keywords: Apple, pear, orchards, ground dwelling, *Coleoptera*, *Staphylinidae*, Hungary.

As part of a larger project (Apple Ecosystem Research), faunistic studies have been done to describe the species composition of arthropod assemblages in apple orchards in Hungary since 1976. Mészáros et al. (1984) examined apple orchards in five localities, Markó et al. (1995) investigated the *Coleoptera* communities in apple and pear orchards in three localities, while Bogyó et al. (1999) presented data on species composition of apple and pear orchard inhabiting *Araneae*. Altogether, more than 2000 animal species were recorded in these three studies. However, there is little information on the structure of ground dwelling beetles, their diversity and abundance in apple and especially in pear orchards (Kádár and Szél, 1989; Kádár and Lővei, 1992; Kutasi et al., 2001).

Staphylinid beetles have been reported from orchard ecosystems by several authors (Galli, 1985; Reede, 1985; Majzlan and Holecová, 1993; Heyer, 1994; Knopp, 1997), but comprehensive fauna lists are rare. Andersen (1991) presented a list of staphylinid beetles occurring in agricultural ecosystems in Norway, namely spring barley, cabbage, carrot, potato, strawberries and grassland fields. The author found 103 000 specimens belonging to 226 staphylinid species. The most frequently found staphylinid species were *Aloconota*

gregaria, *Anotylus rugosus*, *Athena fungi*, *Amischa analis*, *Tachinus signatus* and *Philonthus cognatus*. Levesque and Levesque (1995, 1996) presented a list of staphylinid beetles occurring in raspberry plantations in Canada. The authors presented 81 species and 16,074 individuals (without the species of *Aleocharinae* subfamily). The most dominant were *Gyrophypnus angustatus* and *Tachinus corticinus*. There was a difference in species composition between the old and young plantations. In old raspberry plantations the species *Arpedium cribratum* was common whilst in young raspberry plantation the species *Neohypnus obscurus* and *N. hamatus* were most abundant.

There are frequent reports on Staphylinid beetles from cereal ecosystems in Europe. In wheat, the most abundant species were *Philonthus cognatus*, *Tachyporus hypnorum*, *T. chrysomelinus*, *T. obtusus* and *Stenus biguttatus* (Denis et al., 1990, 1991).

Andersen (2000) recorded the most frequent predator species of *Rhopalosiphum padi* in Norway as *Philonthus cognatus*, *Ph. atratus*, *Ph. ochropus* and *Ph. carbonarius*.

Krooss and Schaefer (1998) studied the effect of different farming systems in cereals, on the occurrence of staphylinid beetles. They found that in winter wheat the dominant species were *T. hypnorum*, *Oxytelus inustus*, *Lesteva longelitrata* and *Ph. fuscipennis*. In nonmanaged farming systems, the most frequent species, were: *Omalium rivulare*, *O. caesum*, *Ph. fuscipennis*, *Ph. rotundikollis*, *Ph. varius*, *Conosoma testeceum*, *Tachinus rufipes* and *Xantholinus linearis*.

The role of staphylinid species in cabbage intercropped with clover was studied by Booij et al. (1997) who found the most important species from the *Philonthus* group.

The parasitoid staphylinid species *Aleochara bilineata* and *A. bipustulata* were recorded in cabbage fields by Ahlstrom-Olsson and Jonasson (1992). In maize the most dominant species were *Neohypnus andinus* and species from the group *Anotylus* (Wardle et al., 1993).

In Hungary several faunistical investigations were carried out in natural ecosystems by Ádám (1996a, b). The author studied staphylinid fauna and identified 334 species from Órség and 509 from Bükk National Park. However, the species composition of staphylinid assemblages in agricultural ecosystems in Hungary have not been studied to date.

Our aims were: (1) to make a thorough survey of the species composition of staphylinid beetles occurring in apple and pear orchards in Hungary, (2) to describe the biodiversity of the staphylinid communities of these orchards and (3) to determine the most widely occurring species in these orchards.

Materials and Methods

The investigations took place in 12 Hungarian orchards, which are located in woodland areas of medium height mountains (Bakonygyirót, Vámosmikola and Pókaszepetk), agricultural lowland environments (Györgyarló, Kecskemét, Tura, Újfehértó, Szentlőrinc) and regularly flooded areas (Szigetcsép). The samples were collected at the following localities: Bakonygyirót (lat. 47° 25' N, long. 17° 48' E, UTM: YN15) (a conventional apple plot), Kecskemét (lat. 54° 40' N, long. 19° 42' E, UTM: CS99) (an abandoned

apple plot), Szigetsép (lat. 47° 16' N, long. 19° E, UTM: CT43) (a conventional apple and a pear plot), Tura (lat. 47° 36' N, long. 19° 36' E, UTM: CT97) (a conventional apple and a pear plot), Újfehértó (lat. 47° 49' N, long. 21° 30' E, UTM: ET59) (a conventional, an abandoned and an "IPM" apple plots), Gyöngytarló (lat. 48° 12' N, long. 21° 30' E, UTM: EU43) (a conventional apple and a pear plot), Szentlőrinc (lat. 46° 3' N, long. 18° E, YM30) (a conventional apple plot), Pókaszeptk (lat. 46° 56' N, long. 16° 58' E, UTM: XM49) (a conventional apple plot), Vámosmikola (lat. 48° N, long. 18° 52' E, UTM: CU31) (a conventional apple plot and near the edge of the orchards) (Fig. 1).

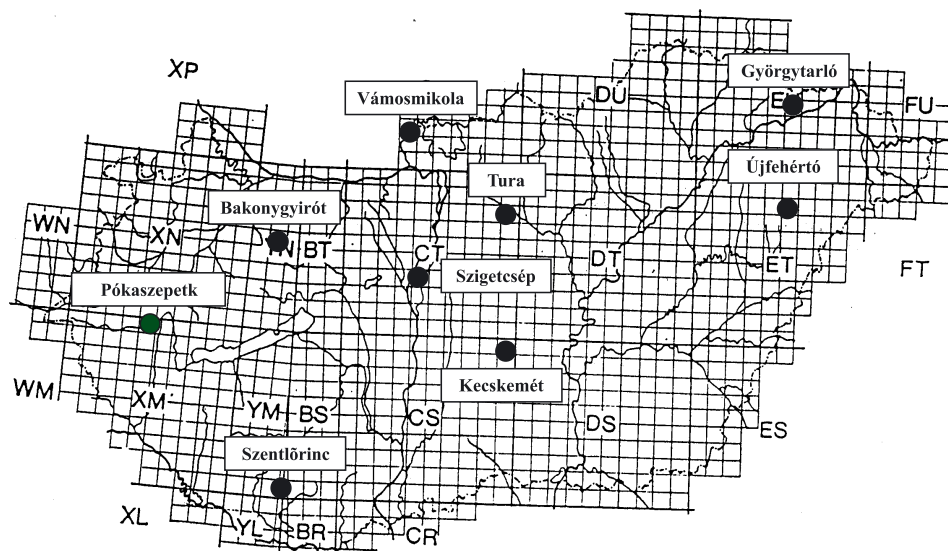


Fig. 1. The UTM map of Hungary with the investigated orchards

The pest management of the orchards was achieved mainly with broad spectrum, organophosphorus insecticides. However, three orchards were investigated in Újfehértó: a conventional, treated with broad spectrum insecticides, another, where some elements of IPM (mostly selective "green" and some "yellow" pesticides) were used, and an abandoned one.

Covered pitfall traps (300 cm³ in size, 8 cm in diameter, half-filled with ethylene glycol 30% solution) were used to collect samples. Ten pitfall traps were placed into the rows in each orchard, except in Újfehértó and Vámosmikola where only six traps were used. Five traps were placed in the centre of the orchards, and five in the inner edges. Samples were collected from April until October in 1998–2001 and traps were emptied monthly.

Table 1 shows the characteristic of every investigated orchard.

The collected staphylinid individuals were identified to the lowest taxonomic level possible using Freude et al. (1964, 1974) and Tóth (1982, 1984).

The most common species in Hungarian orchards were considered either by investigating the total abundance and the frequency (number of localities where a species occurred).

Table 1
The characteristics of the investigated orchards

Locality Environment	Bakonypirót Woodland in mountains	Kecskemét Agricultural lowland	Szigetsép		Tura		Újfehértó		Györgyartló		Szentlőrinc		Pókaszepek		Vámosmikola	
			Flooded forest area	fields	Agricultural lowland	fields, ruderals, orchards	Agricultural lowland	fields, ruderals, orchards	Agr. fields, orchards	apple	pear	Agricultural lowland	fields, orchards	Agricultural lowland	fields, orchards	Woodland in mountains
Neighbouring habitats	Forest (R. pseudacacia)	Agricultural fields, ruderals	Agricultural fields	fields	fields, ruderals, orchards	fields, ruderals, orchards	fields, orchards	fields, orchards	fields, orchards	fields, orchards	fields, orchards	fields, orchards	fields, orchards	fields, orchards	fields, orchards	fields, orchards
Fruits species	apple	apple	apple	pear	apple	pear	apple	apple	apple	pear	apple	apple	apple	apple	apple	apple
Year of planting	1960	1963	1977	1988	1990	1990	1995	1995	1950	1950	1992	1997	1988			
Size of plantation	6 ha	20 ha	5.5 ha	4 ha	118 ha	5 ha	15 ha	20 ha	20 ha	60 ha	1, 2 ha	53 ha	20, 25 ha			
Cultivars	Jt, Bd, S	Jt, S, St	Jt, Jg, G, S	C, V, P, BG	Jt, Ap, Ep	B, D, V	G, Jt, S	Jt	V, B	G, Gl, Jg, I		Jg, Es	I, E			
Planting system	7 × 7 m	5 × 4 m	4.5 × 2.5 m	6 × 4 m	8 × 4 m	7 × 4 m	5 × 2 m 7 × 4 m C	IPM	10 × 10 m	5 × 4 m	4 × 2 m	1.2 × 3.2 m	4 × 1.6 m			
Abandoned		+					+									
Conventionally treated	+		+	+	+	+	+	+	+	+	+	+	+			
IPM								+								
Treatments/year	10–15	–	12–17	12–13	10–12	8–10	15–16	10–12	10–12	10–12	10–14	9	14			
Years of collection	1998–2001	1998–2000	1999– 2001	1999– 2001	1998– 2000	1998– 2000	1999–2001	1998– 2000	1998– 2000	1998– 2000	1998–2000	2001	1999–2000			
Pitfall traps/plot	10	10	10	10	10	10	6	10	10	10	10	10	6			
Soil	Sandy	Sandy	Sandy- loam	Sandy- loam	Sandy- loam	Sandy- loam	Sandy	Clay	Clay	Clay	Clay	Clay	Clay			
Weed management	Mw	NM	Mw	Mw	Cu	Cu	NM, Cu, Cu	Mw	Mw	Mw	Mw	Mw	Mw			

Apple cultivars: Ap – Asztraháni piroos, Bd – Budai domonkos, G – Golden Delicious, Gl – Gloszter, E – Éva, Ep – Egri piroos, Es – Elstar, I – Idared, Jt – Jonathan, Jg – Jonagold, S – Starking, St – Staymared;

Pear cultivars: B – Bosc kobak, BG – Bella di giugno, C – Clapp kedveltje, D – Diel vajkörte, P – Päckhamph's Triumph, V – Vilmos;

Planting system: C – Conventional, IPM – Integrated Pest Management;

Weed management: Cu – Cultivated, M – Mowed, Mw – Mowed, NM – Not Managed

For presenting the frequency-abundance relationship power curve were fitted. In case of the individual orchards the staphylinid species with higher relative abundance than 5% were listed in decreasing order.

The Jaccard index (Krebs, 1989) was used to calculate the similarity in species composition between the apple and pear orchards.

Results and Discussion

Tables 2 and 3 show the species composition of staphylinid communities in the ground level of apple and pear orchards in Hungary. Table 2 shows the staphylinid species occurring in the soil surfaces of apple and pear orchards, with sandy and sandy-loam soil, Table 3 shows staphylinid species occurring in the orchards with clay soil.

Altogether 6099 individuals were collected belonging to 241 staphylinid species. 233 have been identified to species level and 8 staphylinid taxa were determined up to generic level. More than 20% of the Hungarian staphylinid fauna was represented in the sampled orchards. The similarity (Jaccard index) between apple and pear orchard inhabiting *Staphylinidae* was 54%. The species richness in each orchard varied between 23 and 100 species.

In apple orchards with sandy and sandy-loam soil the species with higher relative abundance than 5% in decreasing order were the following:

In Bakonygyirót (conventionally treated plot) 100 species were found, and the most abundant were *Palporus nitidulus*, *Coprochara bipustulata*, *Oligota pumilio*, *Sphenoma abdominale* and *Mocyta orbata*.

In Kecskemét (abandoned plot) 57 species were found, the most abundant were *Drusilla canaliculata*, *Sphenoma abdominale*, *Paraphallus linearis*, *Ocypus nitens* and *Sphenoma togata*.

In Pókaszeptek (conventionally treated plot) a total of 23 species were collected and the most common were *Dinaraea angustula*, *Palporus nitidulus*, *Oligota pumilio* and *Styloxys insecatus*.

In Szigetcsép (conventional plot) 50 species were found and the most widely occurring were *Palporus nitidulus* and *Drusilla canaliculata*.

In Újfehértó a total of 72 species were found (53 in untreated plot, 33 in conventional plot and 22 in IPM plot). Eleven species were recorded in all three plots. The most common species were *Dinaraea angustula*, *Mocyta orbata*, *Coprochara bipustulata* and *Palporus nitidulus* in conventionally treated plot. The number of collected individuals was the lowest in IPM plot and *Styloxys insecatus*, *Coprochara bipustulata* and *Dinaraea angustula* were the most commonly found species. *Omalium caesum*, *Drusilla canaliculata*, *Sphenoma abdominale*, *Podoxya vicina* and *Mocyta orbata* were the species with higher relative abundance than five percent in the untreated plot.

The species with higher abundance in apple orchards with clay soil were the following:

In Györgytarló (conventionally treated plot) 47 species were found and the most common were *Omalium caesum*, *Purrolinus laeticeps*, *Styloxys rugifrons*, *Meneido-phallus roubali* and *Styloxys striatus*.

Table 2
List of staphylinid species occurring on the soil surface of apple and pear orchards with sandy and sandy-loam soil

Species	Bakonypirót		Kecskemét		Szigetcsép		Tura		Újfehértó		Újfehértó	
	CON	apple	AB	apple	CON	apple	CON	apple	AB	apple	CON	apple
<i>Acrolocha minuta</i> (Olivier, 1795)					00							
<i>Alapsodus melanarius</i> (Heer, 1839)			99				00					
<i>Alapsodus winkleri</i> (Bernhauer, 1906)			98, 99				99					
<i>Aleochara bipustulata</i> Gravenhorst, 1802	00											
<i>Aleochara curtula</i> (Goeze, 1777)	00, 01		99		01		00			99, 00		
<i>Aloconota gracilenta</i> (Erichson, 1839)	00									99		
<i>Aloconota gregaria</i> (Erichson, 1839)	98, 00		00		00		00					
<i>Aloconota ruficrus</i> (Stephens, 1832)	01											
<i>Aloconota analis</i> (Gravenhorst, 1802)	00, 01				00						99, 00	
<i>Amischa bifoveolata</i> (Mannerhiem, 1831)												
<i>Amischa decipiens</i> (Sharp, 1869)	00											
<i>Anorylus inustus</i> (Gravenhorst, 1806)	98											
<i>Anorylus nitidulus</i> (Gravenhorst, 1802)	98											
<i>Anorylus rugosus</i> (Fabricius, 1775)	00											
<i>Anorylus sculpturatus</i> (Gravenhorst, 1806)												
<i>Anorylus</i> sp.	99											
<i>Anthobium florale</i> (Paykull, 1789)	00											
<i>Astenus brevelyratus</i> (Coiffait, 1960)	99, 00										99, 00	
<i>Astenus longeliratus</i> Gravenhorst, 1806	00				01							
<i>Atheta aeneicollis</i> (Erichson, 1837)												
<i>Atheta crassicornis</i> (Fabricius, 1792)	00		99, 00				00				00	
<i>Atheta gaggatina</i> (Baudi, 1848)			00									99
<i>Atheta ravilla</i> (Erichson, 1839)												
<i>Atheta sodalis</i> (Erichson, 1837)	98											
<i>Atheta</i> sp.	01											
<i>Atheta triangulum</i> (Kraatz, 1856)	00, 01						00				00	
<i>Atheta trimotata</i> (Kraatz, 1856)												
<i>Atheta xanthopus</i> (Thomson, 1856)	98							98, 00			00	

Table 2 (cont)

Species	Bakonypirót		Kecskemét		Szigetsép		Szigetsép		Tura		Tura		Újfehértó		Újfehértó	
	CON	apple	AB	apple	CON	apple	CON	pear	CON	apple	CON	pear	AB	apple	CON	apple
<i>Bessopora annulata</i> (Mannerheim, 1831)		00														
<i>Bessopora filiforme</i> (L. Redtenbacher, 1849)									99							99
<i>Bessopora haemorrhoum</i> (Mannerheim, 1831)									00				01			
<i>Bolitochara collaris</i> (Paykull, 1800)																
<i>Calodera aethiops</i> (Gravenhorst, 1802)		01														
<i>Carpelimus obesus</i> (Kiessener, 1844)				99, 00				00							99	
<i>Ceranota erythroptera</i> (Gravenhorst, 1806)																
<i>Coprochara bipustulata</i> (Linnaeus, 1761)		98, 99, 00, 01				01		00, 01		98, 00			00, 01		99, 00	99, 01
<i>Craetopycerus cornutus</i> (Gravenhorst, 1802)		00						00								
<i>Craetopycerus nitens</i> (C. R. Sahlberg, 1832)		99, 00				00		00		99						
<i>Craetopycerus rufospinus</i> (Hochhuth, 1851)								00		00						
<i>Craetopycerus spinosus</i> (Erichson, 1840)						01										
<i>Demosoma</i> sp.								00								
<i>Dextiogyia corticina</i> (Erichson, 1837)									99							
<i>Dinaraea angustula</i> (Gyllenhal, 1810)		98, 99, 00, 01				00, 01		00, 01		00			00, 01		99, 00, 01	99, 00, 01
<i>Distichalius flavicornis</i> (Linnaeus, 1758)										98						
<i>Drusilla canaliculata</i> (Fabricius, 1787)																
<i>Enalodroma hepatica</i> (Erichson, 1839)				98, 99, 00		00, 01		00					00, 01		01	99
<i>Falagria caesa</i> Erichson, 1837		00														
<i>Falagria sulcatula</i> (Gravenhorst, 1806)										00						
<i>Gabrius femoralis</i> (Hochhuth, 1851)														00		
<i>Gabrius nigrifolius</i> (Gravenhorst, 1802)		98, 99, 00, 01						00		99						
<i>Gabrius osseticus</i> (Kolenati, 1846)				98, 99, 00				00		00						
<i>Gabrius</i> sp.		01														
<i>Gabrius surveolens</i> (Stephens, 1833)														00		
<i>Gabrius suffragani</i> Jay, 1913		00				00		00		00						
<i>Gefyrobius denigrator</i> (Gravenhorst, 1806)				99												
<i>Goesitba circellaris</i> (Gravenhorst, 1806)				00												99
<i>Hemistenus ludyi</i> (Fauvel, 1886)				98, 99												
<i>Hemistenus ochropus</i> (Kiessener, 1858)				00												

Table 2 (cont)

Species	Bakonypirót		Kecskemét		Szigetcsép		Szigetcsép		Tura		Tura		Újfehértó		Újfehértó	
	CON	apple	AB	apple	CON	apple	CON	pear	CON	apple	CON	pear	CON	apple	CON	apple
<i>Hemistenus</i> sp.																
<i>Hemitropia sordida</i> (Marsham, 1802)	98, 99, 00, 01							00							99	99
<i>Hesperophilus gallicus</i> (Gravenhorst, 1806)								00								
<i>Heterothops dissimilis</i> (Gravenhorst, 1806)	00, 01		99, 00		00, 01		99, 00		98						00	99, 00
<i>Hypomygrus angustatus</i> (Stephens, 1833)	98, 99, 00, 01		98		98, 99, 00, 01		00		99					99	99, 00	99
<i>Hypomygrus fracticornis</i> (O. F. Müller, 1776)	00															
<i>Ilyobates subopacus</i> Palm, 1935	01															
<i>Lathrimaeum atrocephalum</i> (Gyllenhal, 1827)	99						00		00							
<i>Lathrobium boreale</i> Hochhuth, 1851	99															
<i>Lathrobium brunnipes</i> (Fabricius, 1792)	01															
<i>Lathrobium castaneipenne</i> Kolenati, 1846	01															
<i>Lathrobium fovulum</i> Stephens, 1833	00															
<i>Lathrobium</i> sp.	00															
<i>Leptacinus barychrus</i> (Gyllenhal, 1827)																
<i>Leptacinus intermedius</i> Domisthorpe, 1936																
<i>Leptacinus sulcifrons</i> (Stephens, 1833)	98															
<i>Leptobium gracile</i> (Gravenhorst, 1802)																
<i>Leucoparyphus silphoides</i> (Linnaeus, 1767)																
<i>Liogluta crassicornis</i> (Gyllenhal, 1827)	00															
<i>Liogluta longiuscula</i> (Gravenhorst, 1802)	98, 99, 00															
<i>Liogluta oblongiuscula</i> (Sharp, 1869)	00															
<i>Meneidophallus roubali</i> (Coiffait, 1956)	00															
<i>Microdota ganglbaueri</i> (Brundin, 1948)	99															
<i>Microdota pitionii</i> (Scheerpeltz, 1950)	99															
<i>Microsaurus cruentus</i> (Olivier, 1795)																
<i>Microsaurus longicornis</i> (Kraatz, 1857)	98															
<i>Microsaurus ochripennis</i> (Ménétries, 1832)	98															
<i>Mniobates forticornis</i> (Lacordaire, 1835)	98, 00, 01															
<i>Mocytia fungi</i> (Gravenhorst, 1806)	98, 00, 01															
<i>Mocytia negligens</i> (Mulsant et Rey, 1873)																

Table 2 (cont)

Species	Bakonypirót		Kecskemét		Szigetcsép		Szigetcsép		Tura		Tura		Újfehértó		Újfehértó	
	CON	apple	AB	apple	CON	apple	CON	pear	CON	apple	CON	pear	CON	apple	CON	apple
<i>Mocya orbata</i> (Erichson, 1837)	99, 00, 01				00, 01		00, 01		99, 00		99, 00		99, 00, 01		99, 01	
<i>Mycetoporus splendidus</i> (Gravenhorst, 1806)	01				00		00		99, 00		99, 00		00		99, 01	
<i>Mycetoporus forticornis</i> (Fauvel, 1875)	98		99										99			
<i>Mycetoporus longula</i> (Mannerheim, 1831)	01															
<i>Mycetoporus phaedra</i> Gravenhorst, 1802	98															
<i>Mycetoporus piceola</i> (Rey, 1883)	98		00				00, 01		99				99		99	
<i>Mycetoporus</i> sp.																
<i>Mycetota laiticollis</i> (Stephens, 1832)					00						00					
<i>Myopinus elongatulus</i> (Stephens, 1834)															99	
<i>Ocyopus brunnipes</i> (Fabritius, 1781)			98, 99, 00		00		01						00			
<i>Ocyopus nitens</i> (Schränk, 1781)			98, 99, 00		00		00									
<i>Ocyopus olens</i> (O. F. Müller, 1764)					00		01									
<i>Ocyopus ophthalmicus</i> (Scopoli, 1763)			00		00		01									
<i>Ocyopus pedator</i> (Gravenhorst, 1802)			98, 99		01											
<i>Oligota pumilio</i> Kiessenswetter, 1858					01											
<i>Oligota pusillima</i> (Gravenhorst, 1806)	00, 01										00					
<i>Olophrum assimile</i> (Paykull, 1800)																
<i>Omalius caesum</i> Gravenhorst, 1806	98, 00, 01		99, 00		00		00		99, 00		98, 99, 00		99, 00, 01		99, 00, 01	
<i>Omalius cursor</i> (O. F. Müller, 1776)	98, 01		99		00		00		99, 00		98, 99, 00		00		00	
<i>Ontholestes haroldi</i> (Eppelseim, 1884)	99												00		00	
<i>Ontholestes murinus</i> (Linnaeus, 1758)																
<i>Othius laeviusculus</i> (Stephens, 1833)																
<i>Othius punctulatus</i> (Goeze, 1777)					01											
<i>Oxyopoda acuminata</i> (Stephens, 1832)	99, 00, 01								99		00					
<i>Oxyopoda opaca</i> (Gravenhorst, 1802)	01															
<i>Oxyopoda vitata</i> Mörkel, 1842																
<i>Oxytelops tetracariniatus</i> (Black, 1799)	00, 01		99		00		00				00					
<i>Pachnida nigella</i> (Erichson, 1837)	01															
<i>Paederus fuscipes</i> Curtis, 1826					01		00, 01				00		00			
<i>Paederus littoralis</i> Gravenhorst, 1802			98, 99, 00													

Table 2 (cont)

Species	Bakonypirót		Kecskemét		Szigetcsép		Szigetcsép		Tura		Tura		Újfehértó		Újfehértó	
	CON	apple	AB	apple	CON	apple	CON	pear	CON	apple	CON	pear	AB	apple	CON	apple
<i>Pseudocypus vagans</i> (Heer, 1839)						01										
<i>Purrolinus laeticeps</i> (Reitter, 1908)	00		99				00		98				99		99	
<i>Purrolinus tricolor</i> (Fabricius, 1787)	01												00, 01			
<i>Raphirus scintillans</i> (Gravenhorst, 1806)									00							
<i>Que dius fuliginosus</i> (Gravenhorst, 1802)						01			99				00, 01			
<i>Que dius laticollis</i> (Gravenhorst, 1802)													00			
<i>Que dius levicollis</i> Brullé, 1832	98, 00															
<i>Que dius meridiocarpaticus</i> (Gravenhorst, 1806)			99, 00			01	01									
<i>Que dius molochinus</i> (Gravenhorst, 1802)			99			00	00		98							
<i>Rabigus pullus</i> (Nordmann, 1837)							98, 00									
<i>Raphirus limbatooides</i> (Coiffait, 1963)									98							
<i>Raphirus nitipennis</i> Stephens, 1833									00							
<i>Rugilus immunitus</i> Stephens, 1833																
<i>Rugilus orbiculatus</i> (Paykuli, 1789)	99, 00												00			
<i>Rugilus rufipes</i> Germar, 1835			99					99								
<i>Scopaeus bicolor</i> Baudi, 1848	99															
<i>Scopaeus debilis</i> Hochhuth, 1851	99															
<i>Scopaeus minutus</i> Erichson, 1840	00															
<i>Scopaeus</i> sp.									00							
<i>Semiris fusca</i> (Gravenhorst, 1806)						00	00									
<i>Sepedophilus immaculatus</i> (Stephens, 1832)			99													
<i>Sepedophilus marshami</i> (Stephens, 1832)			99, 00				00		99	98, 99, 00		00		99		99
<i>Sepedophilus obtusus</i> (Luzé, 1902)			00													
<i>Sepedophilus pedicularius</i> (Gravenhorst, 1802)																
<i>Sericoderus lateralis</i> (Gyllenhal, 1827)	01															
<i>Sphenonoma abdominale</i> Mannerheim, 1831	98, 99, 00, 01		00			01	00	00	98, 99, 00	00	00	99, 00, 01	00, 01	00, 01		99, 00
<i>Sphenonoma togata</i> (Erichson, 1837)			99					00								
<i>Staphylinus caesareus</i> (Cederhielm, 1798)	00															
<i>Stenus ater</i> Mannerheim, 1831	01															99
<i>Stenus biguttatus</i> (Linnaeus, 1758)	99						00	00								

Table 2 (cont)

Species	Bakonypirót		Kecskemét		Szigetsép		Szigetsép		Tura		Tura		Újfehértó		Újfehértó	
	CON	apple	AB	apple	CON	apple	CON	pear	CON	apple	CON	pear	AB	apple	CON	apple
<i>Stenus clavicornis</i> (Csopoli, 1763)				98, 99, 00												
<i>Stenus juno</i> (Paykull, 1789)	98															
<i>Stenus providus</i> Erichson, 1839							00, 01									
<i>Styloxys insecatus</i> (Gravenhorst, 1806)	98, 00, 01				00		98, 00, 01					98, 00	01			99, 00
<i>Styloxys rugifrons</i> (Hochhuth, 1849)					01		00									
<i>Styloxys striatus</i> (Strom, 1768)	98, 99, 00, 01				00							99, 00		01		
<i>Sutius melanocephalus</i> (Fabricius, 1792)								00								
<i>Tachinus corticinus</i> Gravenhorst, 1802	00			98, 99	00		98, 00, 01					99, 00				
<i>Tachinus fimeitarius</i> Gravenhorst, 1802	00						01					00				
<i>Tachinus rufipes</i> (Linnaeus, 1758)								99								
<i>Tachyporus chrysomelinus</i> (Linnaeus, 1758)																
<i>Tachyporus formosus</i> Matthews, 1838																
<i>Tachyporus hypnorum</i> (Fabricius, 1775)	98, 99, 00, 01			99	00, 01		00, 01		98, 99, 00	99, 01		99, 00		99		
<i>Tachyporus marginellus</i> Stephens, 1832							01									
<i>Tachyporus pusillus</i> Gravenhorst, 1806					01		01									
<i>Tachyporus solutus</i> Erichson, 1839					00		00		99			99, 00				
<i>Trianthus lepidus</i> (Gravenhorst, 1802)							00		99				99			
<i>Tetratopeus rufonitidus</i> (Reitter, 1909)							00									
<i>Trogophiloeus pusillus</i> (Gravenhorst, 1802)	00															
<i>Typholimus laevigatus</i> (Gravenhorst, 1802)	99, 00, 01						00, 01									
<i>Xantholinus coiffaiti</i> (Franz, 1966)	99						00		00							
<i>Xantholinus linearis</i> (Olivier, 1795)	00, 01						00		98, 99, 00	98, 99, 00	98, 99, 00	00		99		
<i>Xantholinus longiventris</i> (Heer, 1839)	99, 00, 01						00, 01		99	99	99	00	00	00, 01		

Explanation: AB – Abandoned

CON – Conventional

IPM – Integrated Pest Management

Table 3

List of Staphylinid species occurring on the soil surface of apple and pear orchards with clay soil

Species	György- tarló CON apple	György- tarló CON pear	Szent- lőrinc CON apple	Póka- szepetk CON apple	Vámos- mikola ED apple	Vámos- mikola CON apple
<i>Alapsodus kirbii</i> (Stephens, 1832)				01	00	
<i>Alapsodus melanarius</i> (Heer, 1839)					99	
<i>Alapsodus morsitans</i> (Rossi, 1790)			00		99, 00	
<i>Aleochara curtula</i> (Goeze, 1777)	99	99				
<i>Aleochara lateralis</i> Heer, 1839				01		
<i>Amischa analis</i> (Gravenhorst, 1802)				01		
<i>Amischa bifoveolata</i> (Mannerhiem, 1831)			00			
<i>Amischa decipiens</i> (Sharp, 1869)				01		
<i>Anotylus inustus</i> (Gravenhorst, 1806)			99, 00			
<i>Anotylus sculpturatus</i> (Gravenhorst, 1806)			99, 00			
<i>Astenus brevelytratus</i> (Coiffait, 1960)			98			
<i>Atheta aeneicollis</i> (Erichson, 1837)	98	98				
<i>Atheta crassicornis</i> (Fabricius, 1792)	99	99	00			99
<i>Atheta triangulum</i> (Kraatz, 1856)	98		00			
<i>Atheta trinotata</i> (Kraatz, 1856)			00			
<i>Baeoglena praecox</i> (Erichson, 1839)			00			
<i>Bolitobius castaneus</i> (Stephens, 1832)			00	01		
<i>Brachida exigua</i> (Heer, 1839)					99	
<i>Coprochara bipustulata</i> (Linnaeus, 1761)			99	01		99
<i>Cordalia obscura</i> (Gravenhorst, 1802)				01		
<i>Dinaraea angustula</i> (Gyllenhal, 1810)	98, 00	98		01	00	99, 00
<i>Drusilla canaliculata</i> (Fabricius, 1787)	98	00	99, 00	01	99, 00	99
<i>Falagria sulcatula</i> (Gravenhorst, 1806)				01		
<i>Falagrioma thoracica</i> (Stephens, 1832)			99, 00			
<i>Gabrius femoralis</i> (Hochhuth, 1851)						99
<i>Gabrius osseticus</i> (Kolenati, 1846)	98, 99				99	99
<i>Gabrius surveolens</i> (Stephens, 1833)	00				00	
<i>Gabrius suffragani</i> Jay, 1913	99		99			
<i>Gauropterus fulgidus</i> (Fabricius, 1787)	00					
<i>Heterothops dissimilis</i> (Gravenhorst, 1806)	98				99	
<i>Heterothops niger</i> Kraatz, 1868			00			
<i>Hyponygrus angustatus</i> (Stephens, 1833)	98, 99	00		01		
<i>Ilyobates nigricollis</i> (Paykull, 1800)	99, 00		99, 00			
<i>Ilyobates subopacus</i> Palm, 1935		98				
<i>Lathrimaemum atrocephalum</i> (Gyllenhal, 1827)			98			99
<i>Lathrobium boreale</i> Hochhuth, 1851	98		00			
<i>Liogluta crassicornis</i> (Gyllenhal, 1827)			99		99	
<i>Medon fuscus</i> (Mannerheim, 1831)	99	98, 99	98			
<i>Meneidophallus roubali</i> (Coiffait, 1956)	98, 99, 00	98, 99, 00			99	
<i>Micropeplus marietti</i> Jaquelin du Val, 1857			98, 99, 00			
<i>Mniobates forticornis</i> (Lacordaire, 1835)		99				
<i>Mocyta fungi</i> (Gravenhorst, 1806)	99	99	98, 99	01		
<i>Mocyta negligens</i> (Mulsant et Rey, 1873)						

Table 3 (cont)

Species	György- tarló CON apple	György- tarló CON pear	Szent- lőrinc CON apple	Póka- szepetk CON apple	Vámos- mikola ED apple	Vámos- mikola CON apple
<i>Mocyta orbata</i> (Erichson, 1837)	99	99	98, 99, 00			
<i>Mycetodrepa alternans</i> (Gravenhorst, 1802)	98, 99					
<i>Mycetoporus splendidus</i> (Gravenhorst, 1806)	98		99			
<i>Mycetoporus clavicornis</i> (Stephens, 1832)			99	01		
<i>Mycetoporus forticornis</i> (Fauvel, 1875)		98				
<i>Mycetoporus nigricollis</i> (Stephens, 1835)			99			
<i>Mycetota laticollis</i> (Stephens, 1832)	99	99	98			
<i>Oligota pumilio</i> Kiessenwetter, 1858			00	01		
<i>Olophrum assimile</i> (Paykull, 1800)					99, 00	99, 00
<i>Omalius caesum</i> Gravenhorst, 1806	98, 99	98, 99, 00	98, 99, 00			99
<i>Omalius cursor</i> (O. F. Müller, 1776)	98, 99	98, 99	00	01		
<i>Ontholestes haroldi</i> (Eppelseim, 1884)					99	
<i>Othius punctulatus</i> (Goeze, 1777)	99		98			
<i>Oxypoda acuminata</i> (Stephens, 1832)	98	98, 00	98, 99, 00			
<i>Oxypoda opaca</i> (Gravenhorst, 1802)			00			
<i>Oxytelops tetracaratus</i> (Black, 1799)		99		01		
<i>Paederus fuscipes</i> Curtis, 1826		00				
<i>Paederus littoralis</i> Gravenhorst, 1802			99		99	99
<i>Paederus schoenherri</i> Czwalina, 1889						00
<i>Palporus nitidulus</i> (Fabritius, 1781)	99, 00	99, 00	99, 00	01	00	
<i>Pella limbata</i> (Paykull, 1789)						99
<i>Philonthus carbonarius</i> (Gravenhorst, 1802)	98, 99, 00	98, 99	99		99, 00	
<i>Philonthus cognatus</i> (Stephens, 1832)	98		99		00	
<i>Philonthus cruentatus</i> (Gmelin, 1790)			99			
<i>Philonthus mannerheimi</i> (Fauvel, 1869)					00	
<i>Philonthus nigrinus</i> (Runde, 1835)	00					
<i>Philonthus ochropus</i> (Gravenhorst, 1802)			00			
<i>Philonthus succicola</i> (Thomson, 1860)	98, 99	98, 99				
<i>Philonthus laminatus</i> (Creutzer, 1799)	98	98				
<i>Plataraea dubiosa</i> (G. Benick, 1935)					99	
<i>Platydracus stercorarius</i> (Olivier, 1795)					99, 00	99, 00
<i>Pseudocypus fulvipennis</i> (Erichson, 1840)					99	
<i>Pseudocypus fuscatus</i> (Gravenhorst, 1802)					99	
<i>Pseudocypus mus</i> (Brullé, 1832)			98, 00		99	99, 00
<i>Pseudocypus penetrans</i> (O. F. Müller, 1776)					00	
<i>Proteinus brachypterus</i> (Fabritius, 1792)	99		99			
<i>Purrolinus laeticeps</i> (Reitter, 1908)	98, 99	98	99, 00		99	99
<i>Purrolinus tricolor</i> (Fabricius, 1787)					00	00
<i>Raphirus scintillans</i> (Gravenhorst, 1806)					99	
<i>Quedius curtipennis</i> Bernhauer, 1908					99	99
<i>Quedius levicollis</i> Brullé, 1832			00			
<i>Quedius meridiocarpaticus</i> (Gravenhorst, 1806)			00			
<i>Raphirus limbatooides</i> (Coiffait, 1963)	98, 00				99	
<i>Raphirus nitipennis</i> Stephens, 1833			99, 00			
<i>Rugilus rufipes</i> Germar, 1835					99	

Table 3 (cont)

Species	György- tarló CON apple	György- tarló CON pear	Szent- lőrinc CON apple	Póka- szepetk CON apple	Vámos- mikola ED apple	Vámos- mikola CON apple
<i>Rugilus similis</i> (Erichson, 1839)		98				
<i>Rugilus subtilis</i> (Erichson, 1840)			99, 00		00	
<i>Sepedophilus marshami</i> (Stephens, 1832)	98	00	98, 99		99, 00	99, 00
<i>Sepedophilus testaceus</i> (Fabritius, 1792)	99					
<i>Sphenoma abdominale</i> Mannerheim, 1831	98, 00	00		01		99
<i>Staphylinus caesareus</i> (Cederhielm, 1798)			00		99, 00	
<i>Staphylinus dimidiaticornis</i> (Gemminge, 1851)			00			
<i>Staphylinus erythropterus</i> (Linnaeus, 1758)		98				
<i>Stenus ater</i> Mannerheim, 1831			99			
<i>Stenus clavicornis</i> (Csopoli, 1763)		98, 99	99			
<i>Styloxys insecatus</i> (Gravenhorst, 1806)		98		01		99, 00
<i>Styloxys rugifrons</i> (Hochhuth, 1849)	98, 99	98, 99				
<i>Styloxys striatus</i> (Strom, 1768)	98, 99, 00	98, 99, 00	99			00
<i>Sunius fallax</i> (Lokay, 1919)			00			
<i>Tachinus corticinus</i> Gravenhorst, 1802		99		01	99, 00	
<i>Tachinus rufipennis</i> Gyllenhal, 1810	98				99	
<i>Tachinus rufipes</i> (Linnaeus, 1758)	99				99	99
<i>Tachyporus chrysomelinus</i> (Linnaeus, 1758)	99	99			99	99, 00
<i>Tachyporus hypnorum</i> (Fabritius, 1775)	98, 99	98, 99	98, 99	01	99	
<i>Tachyporus solutus</i> Erichson, 1839	99		99			
<i>Trogophloeus heidenreichi</i> L. Benick, 1934		99				
<i>Typholinus laevigatus</i> (Gravenhorst, 1802)			99, 00		00	00
<i>Xantholinus linearis</i> (Olivier, 1795)	99	00	99			00
<i>Xantholinus longiventris</i> (Heer, 1839)	00	98	99, 00	01		99

Explication: AB – Abandoned, CON – Conventional; ED – Edge; IPM – Integrated Pest Management

In Szentlőrinc (conventionally treated plot) 62 species were identified and the most abundant were *Ocypus olens*, *Typhlolinus laevigatus*, *Mycetoporus splendidus*, *Purrolinus laeticeps* and *Oligota pusillima*.

In Tura (conventional plot) 46 species were found and the most common were *Paraphallus linearis*, *Sphenoma abdominale*, *Mocyta orbata*, *Omalium caesum* and *Podoxya vicina*.

In Vámosmikola (conventionally treated plot and orchard edge) 55 species were identified (43 near the edge of the orchard, 27 in the middle of the orchard and 15 species were present in both plots). In the centre of the orchard the most common species were *Platydracus stercorarius*, *Sphenoma abdominale*, *Pseudocypus mus*, *Dinaraea angustula* and *Sepedophilus marshami*. The most abundant species in the grassy edge were *Platydracus stercorarius*, *Olophrum assimile* and *Drusilla canaliculata*.

One hundred and twenty-five species were found in pear orchards (79 in Szigetcsép, 39 in Györgytarló and 68 in Tura). The species, occurred most often in Szigetcsép were *Phallolinus longiventris*, *Palporus nitidulus*, *Dinaraea angustula*, *Styloxys insecatus* and *Paraphallus longiventris*.

In Gyöngytarló the species *Omalium caesum*, *Purrolinus laeticeps*, *Dinaraea angustula*, *Aleochara curtula* and *Meneidophallus roubali*, in Tura *Omalium caesum*, *Philonthus mannerheimi*, *Oxytelops tetracarinated* and *Omalium cursor* occurred with higher relative abundance than 5%.

In the total from all the orchards, the most abundant species [with high relative abundance (%)], were as follows: *Omalium caesum* (8.39%), *Drusilla canaliculata* (8.13%), *Sphenoma abdominale* (6.54%), *Palporus nitidulus* (6.12%), *Dinaraea angustula* (4.72%), *Paraphallus linearis* (4.66%), *Mocyta orbata* (3.53%), *Coprochara bipustulata* (3.26%), *Platydracus stercorarius* (2.61%), *Phallolinus longiventris* (2.08%), *Oligota pumilio* (1.98%), *Podoxya vicina* (1.80%), *Tachyporus hypnorum* (1.64%) and *Olophrum assimile* (1.59%). These species represent 57.06% of the total catch.

The most widely occurring staphylinid species, which were found in most of the investigated 16 plots were: *Dinaraea angustula* (14 plots), *Palporus nitidulus* (13 plots), *Tachyporus hypnorum* (13 plots), *Sphenoma abdominale* (13 plots), *Omalium caesum* (12 plots), *Philonthus carbonarius* (12 plots), *Drusilla canaliculata* (12 plots), *Sepedophilus marshami* (12 plots), *Mocyta orbata* (11 plots), *Coprochara bipustulata* (11 plots), *Mocyta fungi* (11 plots), *Hyponygrus angustatus* (11 plots), *Purrolinus laeticeps* (11 plots), *Paraphallus linearis* (10 plots), *Omalium cursor* (10 plots), *Heterothops dissimilis* (10 plots) and *Atheta crassicornis* (10 plots). The frequency-abundance relationship of orchard inhabiting staphylinid species is given in Fig. 2. Species which were found in seven or more plantations were never represented in the total catch with low abundance. On the other hand, all the first eight species in the dominance order occur in more than 10 investigated places.

Comparison of our data with the results from other agroecosystems in Europe indicates that there is large variability between the composition of staphylinids. From the 14

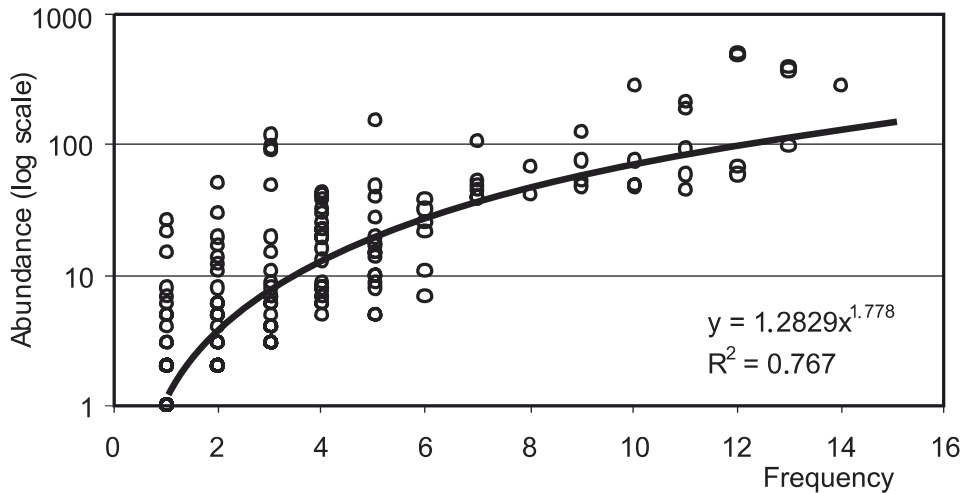


Fig. 2. Relationship between the frequency of occurrence and the total abundance of orchard inhabiting staphylinid species in Hungary

species considered abundant in apple and pear orchards in Hungary only *Omalium caesum*, *Drusilla canaliculata*, *Olophrum assimile* and *Tachyporus hypnorum* were reported as common in European agricultural fields. We can conclude too, that although the dominant staphylinids in Hungarian apple and pear orchards belong to disturbance-resistant species, the staphylinid fauna in these habitats can not be considered uniform. There are significant differences in species composition and especially in dominance order also within Hungary (Kutasi et al., 2001).

Further research is needed to describe the theoretical and practical background of protection and application of Staphylinidae communities in the agroecosystems.

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